

HMMER3 beta test: User's Guide

Biological sequence analysis using profile hidden Markov models

<http://hmmer.org/>
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1 Introduction

This is a user's guide to the HMMER3 test distribution.

It really isn't meant for a new user. I will assume you are already familiar with profile hidden Markov models (profile HMMs) (Krogh et al., 1994; Eddy, 1998; Durbin et al., 1998); with the previous version of HMMER [HMMER2, <http://hmmer.org>]; with other popular biological sequence comparison tools, such as BLAST (Altschul et al., 1997); and with running sequence analysis tools on a UNIX or UNIX-like command line. If this isn't true of you, you should probably not be using the HMMER3 code yet. Instead, you should wait for a later and more stable version, when the user documentation will take less for granted.

Design goals of HMMER3

HMMER3's objective is to combine the power of probabilistic inference with high computational speed. We aim to upgrade some of molecular biology's most important sequence analysis applications to use more powerful statistical inference engines, without sacrificing computational performance.

Specifically, HMMER3 has three main design features that, in combination, distinguish it from previous tools:

Explicit representation of alignment uncertainty. Most sequence alignment analysis tools report only a single best-scoring alignment. However, sequence alignments are uncertain, and the more distantly related sequences are, the more uncertain alignments become. HMMER3 calculates complete posterior alignment ensembles rather than single optimal alignments. Posterior ensembles get used for a variety of useful inferences involving alignment uncertainty. For example, any HMMER3 sequence alignment is accompanied by posterior probability annotation, representing the degree of confidence in each individual aligned residue.

Sequence scores, not alignment scores. Alignment uncertainty has an important impact on the power of sequence database searches. It's precisely the most remote homologs – the most difficult to identify and potentially most interesting sequences – where alignment uncertainty is greatest, and where the statistical approximation inherent in scoring just a single best alignment breaks down the most. To maximize power to discriminate true homologs from nonhomologs in a database search, statistical inference theory says you ought to be scoring sequences by integrating over alignment uncertainty, not just scoring the single best alignment. HMMER3's log-odds scores are sequence scores, not just optimal alignment scores; they are integrated over the posterior alignment ensemble.

Speed. A major limitation of previous profile HMM implementations (including HMMER2) was their slow performance. HMMER3 implements a new heuristic acceleration algorithm. For most queries, it's about as fast as BLAST.

Individually, none of these points is new. As far as alignment ensembles and sequence scores go, pretty much the whole reason why hidden Markov models are so theoretically attractive for sequence analysis is that they are good probabilistic models for explicitly dealing with alignment uncertainty. The SAM profile HMM software from UC Santa Cruz has always used full probabilistic inference (the HMM Forward and Backward algorithms) as opposed to optimal alignment scores (the HMM Viterbi algorithm). HMMER2 had the full HMM inference algorithms available as command-line options, but used Viterbi alignment by default, in part for speed reasons. Calculating alignment ensembles is even more computationally intensive than calculating single optimal alignments.

One reason why it's been hard to deploy sequence scores for practical large-scale use is that we haven't known how to accurately calculate the statistical significance of a log-odds score that's been integrated over alignment uncertainty. Accurate statistical significance estimates are essential when one is trying to discriminate homologs from millions of unrelated sequences in a large sequence database search. The statistical significance of optimal alignment scores can be calculated by Karlin/Altschul statistics (Karlin and

Altschul, 1990, 1993). Karlin/Altschul statistics are one of the most important and fundamental advances introduced by BLAST. However, this theory doesn't apply to integrated log-odds sequence scores (HMM "Forward scores"). The statistical significance (expectation values, or E-values) of HMMER3 sequence scores is determined by using recent theoretical conjectures about the statistical properties of integrated log-odds scores which have been supported by numerical simulation experiments (Eddy, 2008).

And as far as speed goes, there's really nothing new about HMMER3's speed either. Besides Karlin/Altschul statistics, the main reason BLAST has been so useful is that it's so fast. Our design goal in HMMER3 was to achieve rough speed parity between BLAST and more formal and powerful HMM-based methods. The acceleration algorithm in HMMER3 is a new heuristic. It seems likely to be more sensitive than BLAST's heuristics on theoretical grounds. It certainly benchmarks that way in practice (Eddy, 2009, manuscript in preparation). Additionally, it's very well suited to modern hardware architectures. We expect to be able to take good advantage of GPUs and other parallel processing environments in the near future.

What made it into the HMMER3.0 test code

Single sequence queries: new to HMMER3

phmmer	Search a sequence against a sequence database. (BLASTP-like)
jackhmmer	Iteratively search a sequence against a sequence database. (PSIBLAST-like)

Replacements for HMMER2's functionality

hmmbuild	Build a profile HMM from an input multiple alignment.
hmmsearch	Search a profile HMM against a sequence database.
hmmscan	Search a sequence against a profile HMM database.
hmmalign	Make a multiple alignment of many sequences to a common profile HMM.

Other utilities

hmmconvert	Convert profile formats to/from HMMER3 format.
hmmemit	Generate (sample) sequences from a profile HMM.
hmmfetch	Get a profile HMM by name or accession from an HMM database.
hmmcompress	Format an HMM database into a binary format for <code>hmmscan</code> .
hmmstat	Show summary statistics for each profile in an HMM database.

The quadrumvirate of `hmmbuild/hmmsearch/hmmscan/hmmalign` replaces HMMER2's core functionality of `hmmbuild/hmmsearch/hmmpfam/hmmalign` in people's domain analysis and annotation pipelines, for instance using profile databases like Pfam or SMART. These four programs have already been subjected to some serious independent testing by Rob Finn of the Pfam Consortium, who visited Janelia for several months in order to adopt HMMER3 at Pfam, apparently by beating the living tar out of it. I haven't yet fixed all the issues the evil Rob has identified, but I have fixed the showstopping bugs.¹

The `phmmer` and `jackhmmer` programs are new to HMMER3. They searches a single sequence against a sequence database, akin to BLASTP and PSIBLAST, respectively. (Internally, they just produce a profile HMM from the query sequence, then run HMM searches.)

In the Tutorial section, I'll show examples of running each of these programs, using examples in the `tutorial/` subdirectory of the distribution.

¹I think.

What's still missing

Oh, lots. The most egregious lacunae include:

More processor support. One of the attractive features of the MSV algorithm is that it is a very tight and efficient piece of code, which ought to be able to take advantage of recent advances in using massively parallel GPUs (graphics processing units), and other specialized processors such as the Cell processor, or FPGAs. We have prototype work going on in a variety of processors, but none of this is far along as yet. But this work (combined with the parallelization) is partly why we expect to wring significant more speed out of HMMER in the future.

More speed. Even on x86 platforms, HMMER3's acceleration algorithms are still on a nicely sloping bit of their asymptotic optimization curve. I still think I can accelerate the code by another two-fold or so. Additionally, for a small number of HMMs (< 1% of Pfam models), the acceleration core is performing relatively poorly, for reasons I pretty much understand (having to do with biased composition; most of these pesky models are hydrophobic membrane proteins), but which are nontrivial to work around. This'll produce an annoying behavior that some testers are sure to notice: if you look systematically, sometimes you'll see a model that runs at something more like HMMER2 speed, 100x or so slower than an average query. This, needless to say, Will Be Fixed.

DNA sequence comparison. HMMER's search pipeline is somewhat specialized to protein/protein comparison: specifically, the pipeline works by filtering individual sequences, winnowing down to a subset of the sequences in a database that need close attention from the full heavy artillery of Bayesian inference. This strategy doesn't work for long DNA sequences; it doesn't filter the human genome much to say "there's a hit on chromosome 1". The algorithms need to be adapted to identify high-scoring regions of a target sequence, rather than filtering by whole sequence scores. (You can chop a DNA sequence into overlapping windows and HMMER3 would work fine on such a chopped-up database, but that's a disgusting kludge and I don't want to know about it.)

Translated comparisons. We'd of course love to have the HMM equivalents of BLASTX, TBLASTN, and TBLASTX. They'll come.

More sequence input formats. HMMER3 will work fine with FASTA files for unaligned sequences, and Stockholm files for multiple sequence alignments. It has parsers for a handful of other formats (Genbank, EMBL, and Uniprot flatfiles; SELEX format alignments) that we've tested somewhat. It's particularly missing parsers for some widely used alignment formats such as Clustal format, so using HMMER3 on the MSAs produced by many popular multiple alignment programs (MUSCLE or MAFFT for example) is harder than it should be, because it requires a reformat to Stockholm format.

More alignment modes. HMMER3 *only* does local alignment. HMMER2 also could do glocal alignment (align a complete model to a subsequence of the target) and global alignment (align a complete model to a complete target sequence). The E-value statistics of glocal and global alignment remain poorly understood. HMMER3 relies on accurate significance statistics, far more so than HMMER2 did, because HMMER3's acceleration pipeline works by filtering out sequences with poor P-values.

Part of the reason for the test phase is to confirm that these points are just as annoying to you as they are to me, and therefore important to fix asap. Feel free to tell me you want these things even though I already know about them. I also want to find out what glaring problems you find that I'm *not* already losing sleep over. (Really.)

What I hope to accomplish in testing

The core of HMMER3's functionality seems stable to me, but all the stuff wrapped around it – the stuff *you* see, like the applications, command line options, i/o formats – is prototypical and still fluid. The main objective of the test period is for a small number of savvy power users to have the opportunity to give feedback while the user-oriented layers of HMMER3 are still under development – in particular, before its

basic feature set, command line options, and input and output formats get frozen. You might want it to spit out XML, or you like tab-delimited format, or you want this number or that number on such-and-such a line to make it really fit in your analysis pipeline, or you really really need a command line option for slowing the search programs back down to HMMER2 speed so you have more time for coffee². This is the kind of stuff I'd most like to hear now while the code is still fluid.

An obvious corollary of this responsiveness to your feedback is, **don't write any heavy duty output parsers around HMMER3 just yet**. You should expect all the output formats to change, at least slightly, before a public release is finalized.

Of course, since it's test code, I'd like to also hear about bugs: how you manage to break it, or when it produces inconsistent, wrong, or confusing results, or when it doesn't compile or run at all. Some bugs, I already know about, but I'd still like to hear about them just to know you care.

Cryptogenomicon (<http://cryptogenomicon.org/>) is a blog where I'll be talking about issues as they arise in HMMER3, and where you can comment or follow the discussion.

²Actually, this option already exists: `--max`.

2 Installation

Quick installation instructions

Download `hmmmer-3.0b3.tar.gz` from <http://hmmmer.org/>, or directly from <ftp://selab.janelia.org/pub/software/hmmmer3/hmmmer-3.0b3.tar.gz>; `untar`; and change into the newly created directory `hmmmer-3.0b3`:

```
> wget ftp://selab.janelia.org/pub/software/hmmmer3/hmmmer-3.0b3.tar.gz
> tar xf hmmmer-3.0b3.tar.gz
> cd hmmmer-3.0b3
```

The beta test code includes precompiled binaries for x86/Linux platforms. These are in the `binaries` directory. You can just stop here if you like, if you're on a x86/Linux machine and you want to try the programs out without installing them.

To compile new binaries from source, do a standard GNUish build:

```
> ./configure
> make
```

To compile and run a test suite to make sure all is well, you can optionally do:

```
> make check
```

All these tests should pass.

You don't have to install HMMER programs to run them. The newly compiled binaries are now in the `src` directory; you can run them from there. To install the programs and man pages somewhere on your system, do:

```
> make install
```

By default, programs are installed in `/usr/local/bin` and man pages in `/usr/local/man/man1/`. You can change `/usr/local` to any directory you want using the `./configure --prefix` option, as in `./configure --prefix /the/directory/you/want`.

If you have the Intel C compiler `icc`, we strongly recommend that you use it (instead of `gcc`, for example), for performance reasons, by specifying `CC=icc` either in your environment or on the `./configure` command line.

For example, on our systems, we would do:

```
> ./configure CC=icc LDFLAGS=-static --prefix=/usr/local/hmmmer-3.0/
> make
> make check
> make install
```

That's it. You can keep reading if you want to know more about customizing a HMMER3 installation, or you can skip ahead to the next chapter, the tutorial.

System requirements

Operating system: HMMER is designed to run on POSIX-compatible platforms, including UNIX, Linux and MacOS/X. The POSIX standard essentially includes all operating systems except Microsoft Windows.¹

The alpha test code includes precompiled binaries for Linux. These were compiled with the Intel C compiler (`icc`) on an x86_64 Intel platform running Red Hat Enterprise Linux AS4. We believe they should be widely portable to different Linux systems.

We have tested most extensively on Linux, and to a lesser extent on MacOS/X. We aim to be portable to all other POSIX platforms. We currently do not develop or test on Windows.

¹There are add-on products available for making Windows more POSIX-compliant and more compatible with GNU-ish configurations and builds. One such product is Cygwin, <http://www.cygwin.com>, which is freely available. Although we do not test on Windows platforms, we understand HMMER builds fine in a Cygwin environment on Windows.

Processor: HMMER3 depends on vector parallelization methods that are supported on most modern processors. H3 requires either an x86-compatible (IA32/IA64) processor that supports the SSE2 vector instruction set, or a PowerPC processor that supports the AltiVec/VMX instruction set. SSE2 is supported on Intel processors from Pentium 4 on, and AMD processors from K8 (Athlon 64) on; we believe this includes almost all Intel processors since 2000 and AMD processors since 2003. AltiVec/VMX is supported on Motorola G4, IBM G5, and IBM Power6 processors, which we believe includes almost all PowerPC-based desktop systems since 1999 and servers since 2007.

If your platform does not support one of these vector instruction sets, the configure script will revert to an unoptimized implementation called the “dummy” implementation. This implementation is two orders of magnitude slower. It will enable you to see H3’s features on a much wider range of processors, but is not suited for real production work.

We do aim to be portable to all modern processors. The acceleration algorithms are designed to be portable despite their use of specialized SIMD vector instructions. We hope to add support for the Sun SPARC VIS instruction set, for example. We believe that the code will be able to take advantage of GP-GPUs and FPGAs in the future.

Compiler: The source code is C, conforming to POSIX and ANSI C99 standards. It should compile with any ANSI C99 compliant compiler, including the GNU C compiler `gcc`. We test the code using both the `gcc` and `icc` compilers.

If you compile HMMER from source, we strongly recommend using the Intel C compiler `icc` rather than `gcc`. `icc` is free for noncommercial use and heavily discounted for academic use. GNU `gcc` generates HMMER3 code that is significantly slower than `icc` code.² If you find yourself saying, hey, those guys said this program’s supposed to be as fast as BLAST, but it only seems half as fast – odds are you recompiled from source with `gcc`.

Libraries and other installation requirements: HMMER includes a software library called Easel, which it will automatically compile during its installation process. By default, HMMER3 does not require any additional libraries to be installed by you, other than standard ANSI C99 libraries that should already be present on a system that can compile C code. Bundling Easel instead of making it a separate installation requirement is a deliberate design decision to simplify the installation process.³

One of the objectives of the alpha test is to identify portability issues. If HMMER3 fails to compile and/or run fast under a POSIX-compliant OS, on an x86 or PowerPC processor that supports SSE2 or AltiVec/VMX, using an ANSI C99-compliant compiler, please report the problem. If it fails at *all* (with the portable “dummy” implementation) on any POSIX/ANSI C99 compatible platform regardless of processor type, please report the problem.

Multithreaded parallelization for multicores by default

The four search programs support multicore parallelization. This is implemented using POSIX threads. By default, the configure script will identify whether your platform supports POSIX threads (almost all platforms do), and will automatically compile in multithreading support.

If you want to disable multithreading at compile time, recompile from source after giving the `--disable-threads` flag to `./configure`.

²Bjarne Knudsen has identified what’s probably the main reason, and it’s not `gcc`’s fault. We’ll be able to address the speed difference in the near future.

³If you install more than one package that uses the Easel library, it may become an annoyance; you’ll have multiple instantiations of Easel lying around. The Easel API is not yet stable enough to decouple it from the applications that use it, like HMMER and Infernal.

By default, the search programs will use all available cores on your machine. You can control the number of cores each HMMER process will use with the `--cpu <x>` command line option or the `HMMER_NCPU` environment variable. Accordingly, you should observe about a 2x or 4x speedup on dual-core or quad-core machines, relative to previous releases. Even with a single CPU (`--cpu 1`), HMMER will devote a separate execution thread to database input, resulting in significant speedup over serial execution.

If you specify `--cpu 0`, the program will run in serial-only mode, with no threads. This might be useful if you suspect something is awry with the threaded parallel implementation.

MPI parallelization for clusters is optional

The four search programs and `hmmbuild` now also support MPI (Message Passing Interface) parallelization on clusters. To use MPI, you first need to have an MPI library installed, such as OpenMPI (www.open-mpi.org).

MPI support is not enabled by default, and it is not compiled into the precompiled binaries that we supply with HMMER. To enable MPI support at compile time, give the `--enable-mpi` option to the `./configure` command.

To use MPI parallelization, each program that has an MPI-parallel mode has an `--mpi` command line option. This option activates a master/worker parallelization mode. (Without the `--mpi` option, if you run a program under `mpirun` on N nodes, you'll be running N independent duplicate commands, not a single MPI-enabled command. Don't do that.)

It's highly advantageous to get `mpicc` to use the Intel C compiler rather than its default, which is often `gcc`. Different MPI distributions may have different ways of selecting the C compiler and its options. OpenMPI can be controlled by environment variables. For example, in our environment, we currently build HMMER3 for MPI using:

```
> setenv OMPI_CC "icc"
> setenv OMPI_CFLAGS "-O3"
> ./configure --enable-mpi --prefix=/usr/local/hmmer3
> make
```

Using build directories

The installation process from source now supports using separate build directories, using the GNU-standard `VPATH` mechanism. This allows you to maintain separate builds for different processors or with different configuration/compilation options. All you have to do is run the `configure` script from the directory you want to be the root of the build directory. For example:

```
> mkdir my-hmmer-build
> cd my-hmmer-build
> /path/to/hmmer/configure
> make
```

You'd probably only use this feature if you're a developer, maintaining several different builds for testing purposes.

Makefile targets

a11 Builds everything. Same as just saying `make`.

check Runs automated test suites in both HMMER and the Easel library.

clean Removes all files generated by compilation (by `make`). Configuration (files generated by `./configure`) is preserved.

distclean Removes all files generated by configuration (by `./configure`) and by compilation (by `make`).
Note that if you want to make a new configuration (for example, to try an MPI version by `./configure --enable-mpi; make`) you should do a `make distclean` (rather than a `make clean`), to be sure old configuration files aren't used accidentally.

3 Tutorial

Here's a tutorial walk-through of some small projects with HMMER3. This should suffice to get you oriented to a "safe path" through HMMER3 alpha test code that should work as advertised – before you start boldly exploring later-to-be-documented command line options that might or might not be doing anything sensible yet.

Files used in the tutorial

The subdirectory `/tutorial` in the HMMER distribution contains the files used in the tutorial, as well as a number of examples of various file formats that HMMER reads. The important files for the tutorial are:

- globins4.sto** An example alignment of four globin sequences, in Stockholm format. This alignment is a subset of a famous old published structural alignment from Don Bashford (Bashford et al., 1987).
- globins4.hmm** An example profile HMM file, built from `globins4.sto`, in HMMER3 ASCII text format.
- globins4.out** An example `hmmsearch` output file that results from searching the `globins4.hmm` against Uniprot 15.7.
- fn3.sto** An example alignment of 106 fibronectin type III domains. This is the Pfam 22.0 `fn3` seed alignment. It provides an example of a Stockholm format with more complex annotation. We'll also use it for an example of `hmmsearch` analyzing sequences containing multiple domains.
- fn3.hmm** A profile HMM created from `fn3.sto` by `hmmbuild`.
- 7LESS_DROME** A FASTA file containing the sequence of the *Drosophila* Sevenless protein, a receptor tyrosine kinase whose extracellular region is thought to contain seven fibronectin type III domains.
- fn3.out** Output of `hmmsearch fn3.hmm 7LESS_DROME`.
- Pkinase.sto** The Pfam 22.0 Pkinase seed alignment of protein kinase domains.
- minifam** An example HMM flatfile database, containing three models: `globins4`, `fn3`, and `Pkinase`.
- minifam.h3{m,i,f,p}** Binary compressed files corresponding to `minifam`, produced by `hmmcompress`.
- HBB_HUMAN** A FASTA file containing the sequence of human β -hemoglobin, used as an example query for `phmmer` and `jackhmmer`.

Searching a sequence database with a single profile HMM

Step 1: build a profile HMM with hmmbuild

HMMER starts with a multiple sequence alignment file that you provide. Currently HMMER3 only reads Stockholm alignments.¹ The file `tutorial/globins4.sto` is an example of a simple Stockholm file. It looks like this:

¹I'm lying. It can read more. I just don't trust its other parsers yet.

```

# STOCKHOLM 1.0

HBB_HUMAN      . . . . .VHLTPEEKSAVTALWGKV...NVDEVGGEALGRLLVVYPWTQRFFESFGDLSTPDAVMGNPKVKAHGKKVL
HBA_HUMAN      . . . . .VLSPADKTNVKAANGKVA..HAGEYGAEALERMFLSFPTTKTYFPHF.DLS...HGSAQVKGHGKKVA
MYG_PHYCA      . . . . .VLSEGEWQLVHLVWAKVEA..DVAGHGQDILIRLFKSHPETLEKFDKFKHLKTEAEMKASEDLKKHGVTVL
GLB5_PETMA     PIVDTGSVAPLSAAEKTIRSAWAPVYS..TYETSGVDILVKFFTSTPAAQEFFPKFKGLTTADQLKKSADVRWHAERI

HBB_HUMAN      GAFSDGLAHL...D..NLKGTFFATLSELHCCKL..HVDPENFRLLGNVLCVLAHHFGKEFTPPVQAAAYQKVAVAGVANAL
HBA_HUMAN      DALTNAVAHV...D..DMPNALSALSDLHAHKL..RVDPVNFKLLSHCLLVTLAAHLPAEFTPAVHASLDKFLASVSTVL
MYG_PHYCA      TALGAILKK...K.GHHEAELKPLAQSHATKH..KIPKYLEFISEAIIHVLHSRHPGDFGADAQGAMNKALELFRKDI
GLB5_PETMA     NAVNDAVASM..DDTEKMSMKLRDLGSKHAKSF..QVDPQYFKVLAAVIADTVAAG.....DAGFEKLSMVICILL

HBB_HUMAN      AHKYH.....
HBA_HUMAN      TSKYR.....
MYG_PHYCA      AAKYKELGYQG
GLB5_PETMA     RSAY.....
//

```

Most popular alignment formats are similar block-based formats, and can be turned into Stockholm format with a little editing or scripting. Don't forget the `# STOCKHOLM 1.0` line at the start of the alignment, nor the `//` at the end. Stockholm alignments can be concatenated to create an alignment database flatfile containing many alignments.

The `hmmbuild` command builds a profile HMM from an alignment (or HMMs for each of many alignments in a Stockholm file), and saves the HMM(s) in a file. For example, type:

```

> hmmbuild globins4.hmm tutorial/globins4.sto
and you'll see some output that looks like:

```

```

# hmmbuild :: profile HMM construction from multiple sequence alignments
# HMMER 3.0b3 (November 2009); http://hmmer.org/
# Copyright (C) 2009 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# -----
# input alignment file:          globins4.sto
# output HMM file:              globins4.hmm
# -----

# idx name                      nseq  alen  mlen  eff_nseq  re/pos  description
#-----
1    globins4                    4     171   149    0.96    0.589

# CPU time: 0.55u 0.00s 00:00:00.55 Elapsed: 00:00:00.56

```

If your input file had contained more than one alignment, you'd get one line of output for each model. For instance, a single `hmmbuild` command suffices to turn a Pfam seed alignment flatfile (such as `Pfam-A.seed`) into a profile HMM flatfile (such as `Pfam.hmm`).

The information on these lines is almost self-explanatory. The `globins4` alignment consisted of 4 sequences with 171 aligned columns. HMMER turned it into a model of 149 consensus positions, which means it defined 22 gap-containing alignment columns to be insertions relative to consensus. The 4 sequences were only counted as an "effective" total sequence number (`eff_nseq`) of 0.96. The model ended up with a relative entropy per position (`re/pos`; information content) of 0.589 bits.

This output format is rudimentary. HMMER3 knows quite a bit more information about what it's done to build this HMM. Some of this information is likely to be useful to you, the user. As H3 testing and development proceeds, we're likely to expand the amount of data that `hmmbuild` reports.

The new HMM was saved to `globins4.hmm`. If you were to look at this file (and you don't have to – it's intended for HMMER's consumption, not yours), you'd see something like:

```

HMMER3/b [3.0b3 | November 2009]
NAME  globins4
LENG  149
ALPH  amino
RF    no
CS    no
MAP   yes
DATE  Sat Nov 14 10:47:39 2009
NSEQ  4
EFFN  0.964844

```

```

CKSUM 2027839109
STATS LOCAL MSV      -9.9014  0.70957
STATS LOCAL VITERBI -10.7224 0.70957
STATS LOCAL FORWARD -4.1637  0.70957
HMM
  A          C          D          E          F          G          H          I          ...          W          Y
  m->m      m->i      m->d      i->m      i->i      d->m      d->d
COMPO 2.36614 4.52553 2.96662 2.70479 3.20868 3.02051 3.41129 2.90113 ... 4.55399 3.62918
      2.68640 4.42247 2.77497 2.73145 3.46376 2.40504 3.72516 3.29302 ... 4.58499 3.61525
      0.57544 1.78073 1.31293 1.75577 0.18968 0.00000 *
  1    1.70038 4.17733 3.76164 3.36686 3.72281 3.29583 4.27570 2.40482 ... 5.32720 4.10031 9 - -
      2.68618 4.42225 2.77519 2.73123 3.46354 2.40513 3.72494 3.29354 ... 4.58477 3.61503
      0.03156 3.86736 4.58970 0.61958 0.77255 0.34406 1.23405
...
  149  2.92198 5.11574 3.28049 2.65489 4.47826 3.59727 2.51142 3.88373 ... 5.42147 4.18835 165 - -
      2.68634 4.42241 2.77536 2.73098 3.46370 2.40469 3.72511 3.29370 ... 4.58493 3.61418
      0.22163 1.61553 * 1.50361 0.25145 0.00000 *
//

```

The HMMER3 ASCII save file format is defined in Section 4.

If you're used to HMMER2, you may now be expecting to calibrate the model with H2's `hmmcalibrate` program. HMMER3 models no longer need a separate calibration step. We've figured out how to calculate the necessary parameters rapidly, bypassing the need for costly simulation (Eddy, 2008). The determination of the statistical parameters is part of `hmmbuild`. These are the parameter values on the three lines marked `STATS`.

You also may be expecting to need to configure the model's alignment mode, as in HMMER2's `hmmbuild -f` option for building local "fragment search" alignment models, for example. HMMER3's `hmmbuild` does not have these options. `hmmbuild` builds a "core profile", which the search and alignment programs configure as they need to. And at least for the moment, they always configure for local alignment.

Step 2: search the sequence database with `hmmsearch`

Presumably you have a sequence database to search. Here I'll use the Uniprot 15.7 Swissprot FASTA format flatfile (not provided in the tutorial, because of its large size), `uniprot_sprot.fasta`. If you don't have a sequence database handy, run your example search against `tutorial/globins45.fa` instead, which is a FASTA format file containing 45 globin sequences.

`hmmsearch` accepts any FASTA file as input. It also accepts EMBL/Uniprot text format. It will automatically determine what format your file is in; you don't have to say. An example of searching a sequence database with our `globins4.hmm` model would look like:

```
> hmmsearch globins4.hmm uniprot_sprot.fasta > globins4.out
```

Depending on the database you search, the output file `globins4.out` should look more or less like the example of a Uniprot search output provided in `tutorial/globins4.out`.

The first section is the *header* that tells you what program you ran, on what, and with what options:

```

# hmmsearch :: search profile(s) against a sequence database
# HMMER 3.0b3 (November 2009); http://hmmerr.org/
# Copyright (C) 2009 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# -----
# query HMM file:                globins4.hmm
# target sequence database:      uniprot_sprot.fasta
# -----

Query:      globins4 [M=149]
Scores for complete sequences (score includes all domains):

```

The second section is the *sequence top hits* list. It is a list of ranked top hits (sorted by E-value, most significant hit first), formatted in a BLAST-like style:

--- full sequence ---			--- best 1 domain ---			-#dom-				
E-value	score	bias	E-value	score	bias	exp	N	Sequence	Description	
6e-65	222.7	3.2	6.7e-65	222.6	2.2	1.0	1	sp P02185 MYG_PHYCA	Myoglobin OS=Physeter catodon GN=MB PE	
3.1e-63	217.2	0.1	3.4e-63	217.0	0.0	1.0	1	sp P02024 HBB_GORGO	Hemoglobin subunit beta OS=Gorilla gor	

4.5e-63	216.6	0.0	5e-63	216.5	0.0	1.0	1	sp P68871 HBB_HUMAN	Hemoglobin subunit beta OS=Homo sapien
4.5e-63	216.6	0.0	5e-63	216.5	0.0	1.0	1	sp P68872 HBB_PANPA	Hemoglobin subunit beta OS=Pan paniscu
4.5e-63	216.6	0.0	5e-63	216.5	0.0	1.0	1	sp P68873 HBB_PANTR	Hemoglobin subunit beta OS=Pan troglod
6.4e-63	216.1	3.0	7.1e-63	216.0	2.0	1.0	1	sp P02177 MYG_ESCGI	Myoglobin OS=Eschrichtius gibbosus GN=

The last two columns, obviously, are the name of each target sequence and optional description.

The most important number here is the first one, the *sequence E-value*. This is the statistical significance of the match to this sequence: the number of hits we'd expect to score this highly in a database of this size if the database contained only nonhomologous random sequences. The lower the E-value, the more significant the hit.

The E-value is based on the *sequence bit score*, which is the second number. This is the log-odds score for the complete sequence. Some people like to see a bit score instead of an E-value, because the bit score doesn't depend on the size of the sequence database, only on the profile HMM and the target sequence.

The next number, the *bias*, is a correction term for biased sequence composition that's been applied to the sequence bit score.² The only time you really need to pay attention to this value is when it's large, and on the same order of magnitude as the sequence bit score. This might be a sign that the target sequence isn't really a homolog, but merely shares a similar strong biased composition with the query model. The biased composition correction usually works well, but occasionally will not knock down a falsely "significant" nonhomologous hit as far as it should.

The next three numbers are again an E-value, score, and bias, but only for the single best-scoring domain in the sequence, rather than the sum of all its identified domains. The rationale for this isn't apparent in the globin example, because all the globins in this example consist of only a single globin domain. So let's set up a second example, using a model of a single domain that's commonly found in multiple domains in a single sequence. Build a fibronectin type III domain model using the `tutorial/fn3.sto` alignment (this happens to be a Pfam seed alignment; it's a good example of an alignment with complex Stockholm annotation). Then use that model to analyze the sequence `tutorial/7LESS_DROME`, the *Drosophila* Sevenless receptor tyrosine kinase:

```
> hmmbuild fn3.hmm tutorial/fn3.sto
> hmmsearch fn3.hmm tutorial/7LESS_DROME > fn3.out
```

An example of what that output file will look like is provided in `tutorial/fn3.out`. The sequence top hits list says:

--- full sequence ---			--- best 1 domain ---			-#dom-			
E-value	score	bias	E-value	score	bias	exp	N	Sequence	Description
1.9e-57	178.0	0.4	1.2e-16	47.2	0.7	9.4	9	7LESS_DROME	RecName: Full=Protein sevenless;

OK, now let's pick up the explanation where we left off. The total sequence score of 178.0 sums up *all* the fibronectin III domains that were found in the `7LESS_DROME` sequence. The "single best dom" score and E-value are the bit score and E-value as if the target sequence only contained the single best-scoring domain, without this summation.

The idea is that we might be able to detect that a sequence is a member of a multidomain family because it contains multiple weakly-scoring domains, even if no single domain is solidly significant on its own. On the other hand, if the target sequence happened to be a piece of junk consisting of a set of identical internal repeats, and one of those repeats accidentally gives a weak hit to the query model, all the repeats will sum up and the sequence score might look "significant" (which mathematically, alas, is the correct answer: the null hypothesis we're testing against is that the sequence is a *random* sequence of some base composition, and a repetitive sequence isn't random).

So operationally:

- if both E-values are significant ($\ll 1$), the sequence is likely to be homologous to your query.

²The method that HMMER3 uses to compensate for biased composition is unpublished, and different from HMMER2. We will write it up when there's a chance.

- if the full sequence E-value is significant but the single best domain E-value is not, the target sequence is probably a multidomain remote homolog; but be wary, and watch out for the case where it's just a repetitive sequence.

OK, the sharp eyed reader asks, if that's so, then why in the globin4 output (all of which have only a single domain) do the full sequence bit scores and best single domain bit scores not exactly agree? For example, the top ranked hit, MYG_PHYCA (sperm whale myoglobin, if you're curious) has a full sequence score of 222.7 and a single best domain score of 222.6. What's going on? What's going on is that the position and alignment of that domain is uncertain – in this case, only very slightly so, but nonetheless uncertain. The full sequence score is summed over all possible alignments of the globin model to the MYG_PHYCA sequence. When HMMER3 identifies domains, it identifies what it calls an **envelope** bounding where the domain's alignment most probably lies. (More on this later, when we discuss the reported coordinates of domains and alignments in the next section of the output.) The “single best dom” score is calculated after the domain envelope has been defined, and the summation is restricted only to the ensemble of possible alignments that lie within the envelope. The fact that the two scores are slightly different is therefore telling you that there's a small amount of probability (uncertainty) that the domain lies somewhat outside the envelope bounds that HMMER has selected.

The two columns headed #doms are two different estimates of the number of distinct domains that the target sequence contains. The first, the column marked exp, is the *expected* number of domains according to HMMER's statistical model. It's an average, calculated as a weighted marginal sum over all possible alignments. Because it's an average, it isn't necessarily a round integer. The second, the column marked N, is the number of domains that HMMER3's domain postprocessing and annotation pipeline finally decided to identify, annotate, and align in the target sequence. This is the number of alignments that will show up in the domain report later in the output file.

These two numbers should be about the same. Rarely, you might see that they're wildly different, and this would usually be a sign that the target sequence is so highly repetitive that it's confused the H3 domain postprocessors. Such sequences aren't likely to show up as significant homologs to any sensible query in the first place.

The sequence top hits output continues until it runs out of sequences to report. By default, the report includes all sequences with an E-value of 10.0 or less.

Then comes the third output section, which starts with

```
Domain annotation for each sequence (and alignments):
```

Now for each sequence in the top hits list, there will be a section containing a table of where HMMER3 thinks all the domains are, followed by the alignment inferred for each domain. Let's use the fn3 vs. 7LESS_DROME example, because it contains lots of domains, and is more interesting in this respect than the globin4 output. The domain table for 7LESS_DROME looks like:

```
>> 7LESS_DROME RecName: Full=Protein sevenless; EC=2.7.10.1;
# score bias c-Evalue i-Evalue hmmfrom hmm to alifrom ali to envfrom env to acc
---
1 ? -1.3 0.0 0.17 0.17 61 74 .. 396 409 .. 395 411 .. 0.85
2 ! 40.7 0.0 1.3e-14 1.3e-14 2 84 .. 439 520 .. 437 521 .. 0.95
3 ! 14.4 0.0 2e-06 2e-06 13 85 .. 836 913 .. 826 914 .. 0.73
4 ! 5.1 0.0 0.0016 0.0016 10 36 .. 1209 1235 .. 1203 1259 .. 0.82
5 ! 24.3 0.0 1.7e-09 1.7e-09 14 80 .. 1313 1380 .. 1304 1386 .. 0.82
6 ? 0.0 0.0 0.063 0.063 58 72 .. 1754 1768 .. 1739 1769 .. 0.89
7 ! 47.2 0.7 1.2e-16 1.2e-16 1 85 [. 1799 1890 .. 1799 1891 .. 0.91
8 ! 17.8 0.0 1.8e-07 1.8e-07 6 74 .. 1904 1966 .. 1901 1976 .. 0.90
9 ! 12.8 0.0 6.6e-06 6.6e-06 1 86 [] 1993 2107 .. 1993 2107 .. 0.89
```

Domains are reported in the order they appear in the sequence, not in order of their significance.

The ! or ? symbol indicates whether this domain does or does not satisfy both per-sequence and per-domain inclusion thresholds. Inclusion thresholds are used to determine what matches should be considered to be “true”, as opposed to reporting thresholds that determine what matches will be reported (often including the top of the noise, so you can see what interesting sequences might be getting tickled by

your search). By default, inclusion thresholds usually require a per-sequence E value of 0.01 or less and a per-domain conditional E-value of 0.01 or less (except jackhmmer, which requires a more stringent 0.001 for both), and reporting E-value thresholds are set to 10.0.

The bit score and bias values are as described above for sequence scores, but are the score of just one domain's envelope.

The first of the two E-values is the **conditional E-value**. This is an odd number, and it's not even clear we're going to keep it. Pay attention to what it means! It is an attempt to measure the statistical significance of each domain, *given that we've already decided that the target sequence is a true homolog*. It is the expected number of *additional* domains we'd find with a domain score this big in the set of sequences reported in the top hits list, if those sequences consisted only of random nonhomologous sequence outside the region that sufficed to define them as homologs.

The second number is the **independent E-value**: the significance of the sequence in the *whole* database search, if this were the only domain we had identified. It's exactly the same as the "best 1 domain" E-value in the sequence top hits list.

The difference between the two E-values is not apparent in the `7LESS_DROME` example because in both cases, the size of the search space is 1 sequence. There's a single sequence in the target sequence database (that's the size of the search space that the independent/best single domain E-value depends on). There's one sequence reported as a putative homolog in the sequence top hits list (that's the size of the search space that the conditional E-value depends on). A better example is to see what happens when we search Uniprot (15.7 contains 497293 sequences) with the `fn3` model:

```
> hmmsearch fn3.hmm uniprot_sprot.fasta
```

(If you don't have Uniprot and can't run a command like this, don't worry about it - I'll show the relevant bits here.) Now the domain report for `7LESS_DROME` looks like:

#	score	bias	c-Value	i-Value	hmmfrom	hmm to	alifrom	ali to	envfrom	env to	acc
1 !	40.7	0.0	9.1e-12	6.4e-09	2	84 ..	439	520 ..	437	521 ..	0.95
2 !	14.4	0.0	0.0014	1	13	85 ..	836	913 ..	826	914 ..	0.73
3 ?	5.1	0.0	1.1	7.9e+02	10	36 ..	1209	1235 ..	1203	1259 ..	0.82
4 !	24.3	0.0	1.2e-06	0.00084	14	80 ..	1313	1380 ..	1304	1386 ..	0.82
5 !	47.2	0.7	8.3e-14	5.8e-11	1	85 [.	1799	1890 ..	1799	1891 ..	0.91
6 !	17.8	0.0	0.00013	0.091	6	74 ..	1904	1966 ..	1901	1976 ..	0.90
7 !	12.8	0.0	0.0047	3.3	1	86 [.]	1993	2107 ..	1993	2107 ..	0.89

Notice that *almost* everything's the same (it's the same target sequence, after all) *except* for what happens with E-values. The independent E-value is calculated assuming a search space of all 497293 sequences. For example, look at the highest scoring domain (domain 5 here; domain 7 above). When we only looked at a single sequence, its score of 47.2 bits has an E-value of 5.8e-11. When we search a database of 497293 sequences, a hit scoring 47.2 bits would be expected to happen 497293 times as often: $1.2e-16 \times 497293 = 5.97e-11$ (it's showing 5.8e-11 because of roundoff issues; the 1.2e-16 in fact isn't exactly 1.2e-16 inside HMMER). In this Uniprot search, 711 sequences were reported in the top hits list (with E-values ≤ 10). If we were to assume that all 711 are true homologs, x out the domain(s) that made us think that, and then went looking for *additional* domains in those 711 sequences, we'd be searching a smaller database of 711 sequences: the expected number of times we'd see a hit of 47.2 bits or better is now $1.2e-16 \times 711 = 8.3e-14$. That's where the conditional E-value (c-Value) is coming from.

Notice that a couple of domains disappeared in the Uniprot search, because now, in this larger search space size, they're not significant. Domain 1 (the one with the score of -1.3 bits) got a conditional E-value of $0.17 \times 711 = 121$, and domain 6 (with a bit score of 0.0) got a c-Value of $0.063 \times 711 = 45$. These fail the default reporting threshold of 10.0. The domain with a score of 5.1 bits also shifted from being above to below the default inclusion thresholds, so now it's marked with a ? instead of a !.

Operationally:

- If the independent E-value is significant ($\ll 1$), that means that even this single domain *by itself* is such a strong hit that it suffices to identify the sequence as a significant homolog with respect to the size of the entire original database search. You can be confident that this is a homologous domain.

- Once there's one or more high-scoring domains in the sequence already, sufficient to decide that the sequence contains homologs of your query, you can look (with some caution) at the conditional E-value to decide the statistical significance of additional weak-scoring domains.

In the Uniprot output, for example, I'd be pretty sure of four of the domains (1, 4, 5, and maybe 6), each of which has a strong enough independent E-value to declare `7LESS_DROME` to be an fnIII-domain-containing protein. Domains 2 and 7 wouldn't be significant if they were all I saw in the sequence, but once I decide that `7LESS_DROME` contains fn3 domains on the basis of the other hits, their conditional E-values indicate that they are probably also fn3 domains too. Domain 3 is too weak to be sure of, from this search alone, but would be something to pay attention to.

The next four columns give the endpoints of the reported local alignment with respect to both the query model ("hmm from" and "hmm to") and the target sequence ("ali from" and "ali to").

It's not immediately easy to tell from the "to" coordinate whether the alignment ended internally in the query or target, versus ran all the way (as in a full-length global alignment) to the end(s). To make this more readily apparent, with each pair of query and target endpoint coordinates, there's also a little symbology. `..` meaning both ends of the alignment ended internally, and `[]` means both ends of the alignment were full-length flush to the ends of the query or target, and `[.` and `.]` mean only the left or right end was flush/full length.

The next two columns ("env from" and "env to") define the *envelope* of the domain's location on the target sequence. The envelope is almost always a little wider than what HMMER chooses to show as a reasonably confident alignment. As mentioned earlier, the envelope represents a subsequence that encompasses most of the posterior probability for a given homologous domain, even if precise endpoints are only fuzzily inferrable. You'll notice that for higher scoring domains, the coordinates of the envelope and the inferred alignment will tend to be in tighter agreement, corresponding to sharper posterior probability defining the location of the homologous region.

Operationally, I would use the envelope coordinates to annotate domain locations on target sequences, not the alignment coordinates. However, be aware that when two weaker-scoring domains are close to each other, envelope coordinates can and will overlap, corresponding to the overlapping uncertainty of where one domain ends and another begins. In contrast, alignment coordinates generally do not overlap (though there are cases where even they will overlap³).

The last column is the average posterior probability of the aligned target sequence residues; effectively, the expected accuracy per residue of the alignment.

For comparison, current Uniprot consensus annotation of Sevenless shows seven domains:

FT	DOMAIN	311	431	Fibronectin type-III 1.
FT	DOMAIN	436	528	Fibronectin type-III 2.
FT	DOMAIN	822	921	Fibronectin type-III 3.
FT	DOMAIN	1298	1392	Fibronectin type-III 4.
FT	DOMAIN	1680	1794	Fibronectin type-III 5.
FT	DOMAIN	1797	1897	Fibronectin type-III 6.
FT	DOMAIN	1898	1988	Fibronectin type-III 7.

These domains are a pretty tough case to call, actually. HMMER fails to see anything significant overlapping two of these domains (311-431 and 1680-1794) in the Uniprot search, though it sees a smidgen of them when `7LESS_DROME` alone is the target. HMMER3 sees two new domains (1205-1235 and 1993-2098) that Uniprot currently doesn't annotate, but these are pretty plausible domains (given that the extracellular domain of Sevenless is pretty much just a big array of ~100aa fibronectin repeats).

Under the domain table, an "optimal posterior accuracy" alignment (Holmes, 1998) is computed within each domain's envelope, and displayed. For example, (skipping domain 1 because it's weak and unconvincing), fibronectin III domain 2 in your `7LESS_DROME` output is shown as:

```
== domain 2      score: 40.7 bits;   conditional E-value: 1.3e-14
---CEEEEEECTTEEEEE--S..SS--SEEEEEETTTCGCEEEEEETTSEEEEE--TT-EEEEEEEEETTEE.E CS
```

³Not to mention one (mercifully rare) bug/artifact that I'm betting is so unusual that testers don't even see an example of it – but we'll see.

```

fn3 2 saPenlsvsevtstsltlswspkdgppitgYeveyqekgegewqevtvprrtttsvtltgLepgteYefrVqavngagegp 84
saP ++ + ++ l ++W p + +gpi+gY+++++++ + e+ vp+ s+ +++L+gt+Y++ + +n++gegp
7LESS_DROME 439 SAPVIEHLMGLDDSHLAVHWHPRFTNGPIEGYRLRLSSSEGNA-TSEQLVPAGRGSYIFSQLQAGTNYTLALSMINKQGE 520
78999999999*****9998.*****9997 PP

```

The initial header line starts with a == as a little handle for a parsing script to grab hold of. The rest of that line, we'll probably put more information on eventually.

If the model had any consensus structure or reference line annotation that it inherited from your multiple alignment (#=GC SS_cons, #=GC RF annotation in Stockholm files), that information is simply regurgitated as CS or RF annotation lines here. The fn3 model had a consensus structure annotation line.

The line starting with fn3 is the consensus of the query model. Capital letters represent the most conserved (high information content) positions. Dots (.) in this line indicate insertions in the target sequence with respect to the model.

The midline indicates matches between the query model and target sequence. A + indicates positive score, which can be interpreted as “conservative substitution”, with respect to what the model expects at that position.

The line starting with 7LESS_DROME is the target sequence. Dashes (-) in this line indicate deletions in the target sequence with respect to the model.

The bottom line is new to HMMER3. This represents the posterior probability (essentially the expected accuracy) of each aligned residue. A 0 means 0-5%, 1 means 5-15%, and so on; 9 means 85-95%, and a * means 95-100% posterior probability. You can use these posterior probabilities to decide which parts of the alignment are well-determined or not. You'll often observe, for example, that expected alignment accuracy degrades around locations of insertion and deletion, which you'd intuitively expect.

You'll also see expected alignment accuracy degrade at the ends of an alignment – this is because “alignment accuracy” posterior probabilities currently not only includes whether the residue is aligned to one model position versus others, but also confounded with whether a residue should be considered to be homologous (aligned to the model somewhere) versus not homologous at all.⁴

These domain table and per-domain alignment reports for each sequence then continue, for each sequence that was in the per-sequence top hits list.

Finally, at the bottom of the file, you'll see some summary statistics. For example, at the bottom of the globins search output, you'll find something like:

```

Internal pipeline statistics summary:
-----
Query model(s):                1 (149 nodes)
Target sequences:              497293 (175274722 residues)
Passed MSV filter:             19416 (0.0390434); expected 9945.9 (0.02)
Passed bias filter:            15929 (0.0320314); expected 9945.9 (0.02)
Passed Vit filter:             2207 (0.00443803); expected 497.3 (0.001)
Passed Fwd filter:             1076 (0.00216371); expected 5.0 (1e-05)
Initial search space (Z):      497293 [actual number of targets]
Domain search space (domZ):    1075 [number of targets reported over threshold]
# CPU time: 7.08u 0.09s 00:00:07.17 Elapsed: 00:00:02.95
# Mc/sec: 8852.86
//

```

This gives you some idea of what's going on in HMMER3's acceleration pipeline. You've got one query HMM, and the database has 497,293 target sequences. Each sequence goes through a gauntlet of three scoring algorithms called MSV, Viterbi, and Forward, in order of increasing sensitivity and increasing computational requirement.

MSV (the “Multi ungapped Segment Viterbi” algorithm) is the new algorithm in HMMER3. It essentially calculates the HMM equivalent of BLAST's sum score – an optimal sum of ungapped high-scoring alignment segments. Unlike BLAST, it does this calculation directly, without BLAST's word hit or hit extension step, using a SIMD vector-parallel algorithm. By default, HMMER3 is configured to allow sequences with a P-value of ≤ 0.02 through the MSV score filter (thus, if the database contained no homologs and P-values

⁴It may make more sense to condition the posterior probabilities on the assumption that the residue is indeed homologous: given that, how likely is it that I've got it correctly aligned.

were accurately calculated, the highest scoring 2% of the sequences will pass the filter). Here, about 4% of the database got through the MSV filter.

A quick check is then done to see if the target sequence is “obviously” so biased in its composition that it’s unlikely to be a true homolog. This is called the “bias filter”. If you don’t like it (it can occasionally be overaggressive) you can shut it off with the `--nobias` option. Here, 15929 sequences pass through the bias filter.

The Viterbi filter then calculates a gapped optimal alignment score. This is a bit more sensitive than the MSV score, but the Viterbi filter is about four-fold slower than MSV. By default, HMMER3 lets sequences with a P-value of ≤ 0.001 through this stage. Here (because there’s a little over a thousand true globin homologs in this database), much more than that gets through - 2207 sequences.

Then the full Forward score is calculated, which sums over all possible alignments of the profile to the target sequence. The default allows sequences with a P-value of $\leq 10^{-5}\%$ through; 1076 sequences passed.

All sequences that make it through the three filters are then subjected to a full probabilistic analysis using the HMM Forward/Backward algorithms, first to identify domains and assign domain envelopes; then within each individual domain envelope, Forward/Backward calculations are done to determine posterior probabilities for each aligned residue, followed by optimal accuracy alignment. The results of this step are what you finally see on the output.

Recall the difference between conditional and independent E-values, with their two different search space sizes. These search space sizes are reported in the statistics summary.

Finally, it reports the speed of the search in units of Mc/sec (million dynamic programming cells per second), the CPU time, and the elapsed time. This search took about 2.95 seconds of elapsed (wall clock time) (running with `--cpu 2` on two cores). That’s in the same ballpark as BLAST, depending on which BLAST you compare to. On the same machine, also running dual-core, NCBI BLAST with one of these globin sequences took 3.8 seconds, and WU-BLAST took 7.4 seconds.

Searching a profile HMM database with a query sequence

The `hmmscan` program is for annotating all the different known/detectable domains in a given sequence. It takes a single query sequence and an HMM database as input. The HMM database might be Pfam, SMART, or TIGRFams, for example, or another collection of your choice.

Step 1: create an HMM database flatfile

An HMM “database” flatfile is simply a concatenation of individual HMM files. To create a database flatfile, you can either build individual HMM files and concatenate them, or you can concatenate Stockholm alignments and use `hmmbuild` to build an HMM database of all of them in one command.

Let’s create a tiny database called `minifam` containing models of globin, fn3, and Pkinase (protein kinase) domains by concatenating model files:

```
> hmmbuild globins4.hmm tutorial/globins4.sto
> hmmbuild fn3.hmm tutorial/fn3.sto
> hmmbuild Pkinase.hmm tutorial/Pkinase.sto
> cat globins4.hmm fn3.hmm Pkinase.hmm > minifam
```

We’ll use `minifam` for our examples in just a bit, but first a few words on other ways to build HMM databases, especially big ones. The file `tutorials/minifam` is the same thing, if you want to just use that.

Alternatively, you can concatenate Stockholm alignment files together (as Pfam does in its big `Pfam-A.seed` and `Pfam-A.full` flatfiles) and use `hmmbuild` to build HMMs for all the alignments at once. This won’t work properly for our tutorial alignments, because the `globins4.sto` alignment doesn’t have an `#=GF ID` annotation line giving a name to the globins4 alignment, so `hmmbuild` wouldn’t know how to name it correctly. To build a multi-model database from a multi-MSA flatfile, the alignments have to be in Stockholm format (no

other MSA format that I'm aware of supports having more than one alignment per file), and each alignment must have a name on a `#=GF ID` line.

But if you happen to have a Pfam seed alignment flatfile `Pfam-A.seed` around, an example command would be:

```
> hmmbuild Pfam-A.hmm Pfam-A.seed
```

This would take about two or three hours to build all 10,000 models or so in Pfam. To speed the database construction process up, `hmmbuild` supports MPI parallelization.

As far as HMMER's concerned, all you have to do is add `--mpi` to the command line for `hmmbuild`, assuming you've compiled support for MPI into it (see the installation instructions). You'll also need to know how to invoke an MPI job in your particular environment, with your job scheduler and MPI distribution. We can't really help you with this – different sites have different cluster environments.

With our scheduler (SGE, the Sun Grid Engine) and our MPI distro (OpenMPI), an example incantation for building `Pfam.hmm` from `Pfam-A.seed` is:

```
> qsub -N hmmbuild -j y -o errors.out -b y -cwd -V -pe openmpi 128
'mpirun -mca btl self,openib,sm -np 128 ./hmmbuild --mpi Pfam.hmm Pfam-A.seed > hmmbuild.out'
```

This reduces the time to build all of Pfam to about 40 seconds.

Step 2: compress and index the flatfile with hmmpress

The `hmmscan` program has to read a lot of profile HMMs in a hurry, and HMMER's ASCII flatfiles are bulky. To accelerate this, `hmmscan` uses binary compression and indexing of the flatfiles. To use `hmmscan`, you must first compress and index your HMM database with the `hmmpress` program:

```
> hmmpress minifam
```

This will quickly produce:

```
Working... done.
Pressed and indexed 3 HMMs (3 names and 2 accessions).
Models pressed into binary file: minifam.h3m
SSI index for binary model file: minifam.h3i
Profiles (MSV part) pressed into: minifam.h3f
Profiles (remainder) pressed into: minifam.h3p
```

and you'll see these four new binary files in the directory.

The `tutorial/minifam` example has already been pressed, so there are example binary files `tutorial/minifam.h3{m,i,f,p}` included in the tutorial.

Their format is "proprietary", which is an open source term of art that means both "I haven't found time to document them yet" and "I still might decide to change them arbitrarily without telling you".

Step 3: search the HMM database with hmmscan

Now we can analyze sequences using our HMM database and `hmmscan`.

For example, the receptor tyrosine kinase `7LESS_DROME` not only has all those fibronectin type III domains on its extracellular face, it's got a protein kinase domain on its intracellular face. Our `minifam` database has models of both `fn3` and `Pkinase`, as well as the unrelated `globins4` model. So what happens when we scan the `7LESS_DROME` sequence:

```
> hmmscan minifam tutorial/7LESS_DROME
```

The header and the first section of the output will look like:

```
# hmmscan :: search sequence(s) against a profile database
# HMMER 3.0b3 (November 2009); http://hmmer.org/
# Copyright (C) 2009 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# -----
# query sequence file:          7LESS_DROME
# target HMM database:         minifam
# per-seq hits tabular output:  7LESS.tbl
# per-dom hits tabular output:  7LESS.domtbl
```

```

# -----
Query:          7LESS_DROME [L=2554]
Accession:      P13368
Description:    RecName: Full=Protein sevenless;          EC=2.7.10.1;
Scores for complete sequence (score includes all domains):
--- full sequence ---   --- best 1 domain ---   -#dom-
E-value  score  bias    E-value  score  bias    exp  N  Model  Description
-----
5.6e-57  178.0  0.4    3.5e-16  47.2  0.7    9.4  9  fn3    Fibronectin type III domain
1.1e-43  137.2  0.0    1.7e-43  136.5  0.0    1.3  1  Pkinase Protein kinase domain

```

The output fields are in the same order and have the same meaning as in `hmmsearch`'s output.

The size of the search space for `hmmsearch` is the number of models in the HMM database (here, 3; for a Pfam search, on the order of 10000). In `hmmsearch`, the size of the search space is the number of sequences in the sequence database. This means that E-values may differ even for the same individual profile vs. sequence comparison, depending on how you do the search.

For domain, there then follows a domain table and alignment output, just as in `hmmsearch`. The `fn3` annotation, for example, looks like:

```

Domain and alignment annotation for each model:
>> fn3 Fibronectin type III domain
#   score  bias  c-Evalue  i-Evalue  hmmfrom  hmm to  alifrom  ali to  envfrom  env to  acc
---
1 ?   -1.3  0.0    0.33     0.5      61      74 ..   396     409 ..   395     411 ..  0.85
2 !   40.7  0.0    2.6e-14  3.8e-14   2       84 ..   439     520 ..   437     521 ..  0.95
3 !   14.4  0.0    4.1e-06  6.1e-06   13      85 ..   836     913 ..   826     914 ..  0.73
4 !    5.1  0.0    0.0032   0.0048   10      36 ..  1209    1235 ..  1203    1259 ..  0.82
5 !   24.3  0.0    3.4e-09  5e-09    14      80 ..  1313    1380 ..  1304    1386 ..  0.82
6 ?    0.0  0.0     0.13     0.19    58      72 ..  1754    1768 ..  1739    1769 ..  0.89
7 !   47.2  0.7    2.3e-16  3.5e-16   1       85 [..  1799    1890 ..  1799    1891 ..  0.91
8 !   17.8  0.0    3.7e-07  5.5e-07   6       74 ..  1904    1966 ..  1901    1976 ..  0.90
9 !   12.8  0.0    1.3e-05  2e-05     1       86 []  1993    2107 ..  1993    2107 ..  0.89

```

and an example alignment (of that second domain again):

```

== domain 2      score: 40.7 bits; conditional E-value: 2.6e-14
---CEEEEEEECTTEEEEEEE--S..SS--SEEEEEEEETTCCGCEEEEEETTSEEEEEES--TT-EEEEEEEEETTEE.E CS
fn3 2 saPenlsvsevtstsltlSwsppkdgggpigtYeveyqekgegewevtvprtttsvtltgLepgteYefrVqavngagegp 84
saP ++ + ++ l ++W p + +gpitgY+++++++ + e+ vp+ s+ +++L++gt+Y++ + +n++gegp
7LESS_DROME 439 SAPVIEHLMGLDDSHLAVHWHPRFTNGPIEGYRLRLSSSEGNA-TSEQLVPAGRGSYIFSQLQAGTNYTLALSMINKQGE 520
789999999999*****9998*****9997 PP

```

You'd think that except for the E-values (which depend on database search space sizes), you should get exactly the same scores, domain number, domain coordinates, and alignment every time you do a search of the same HMM against the same sequence. And this is actually the case – but in fact, it's actually not so obvious this should be so, and HMMER is going out of its way to make it so. HMMER uses stochastic sampling algorithms to infer some parameters, and also to infer the exact domain number and domain boundaries in certain difficult cases. If HMMER ran its stochastic samples "properly", it would see different samples every time you ran a program, and all of you would complain to me that HMMER was weird and buggy because it gave different answers on the same problem. To suppress run-to-run variation, HMMER seeds its random number generator(s) identically every time you do a sequence comparison. If you're an expert, and you really want to see the proper stochastic variation that results from any sampling algorithms that got run, you can pass a command-line argument of `--seed 0` to programs that have this property (`hmmbuild` and the four search programs).

Creating multiple alignments with `hmmalign`

The file `tutorial/globins45.fa` is a FASTA file containing 45 unaligned globin sequences. To align all of these to the `globins4` model and make a multiple sequence alignment:

```
> hmmalign globins4.hmm tutorial/globins45.fa
```

The output of this is a Stockholm format multiple alignment file. The first few lines of it look like:

```

# STOCKHOLM 1.0

MYG_ESCGI      .-VLSDAEWQLVLNIWAKVEADVAGHGQDILIRLFGKHPETLEKFDKFKH
#=GR MYG_ESCGI PP  .69*****
MYG_HORSE      g--LSDGEWQQVLNVWGKVEAD IAGHGQEVLI RLF TGH PETLEKFDKFKH
#=GR MYG_HORSE PP  8..89*****
MYG_PROGU      g--LSDGEWQLVLNVWGKVEGDL SGHGQEVLI RLF KGH PETLEKFDKFKH
#=GR MYG_PROGU PP  8..89*****
MYG_SAISC      g--LSDGEWQLVLNIWGKVEADIP SHGQEVLI SLFKGHPETLEKFDKFKH
#=GR MYG_SAISC PP  8..89*****
MYG_LYCPI      g--LSDGEWQIVLNIWGKVETDL AGHGQEVLI RLF KNH PETLDKFDKFKH
#=GR MYG_LYCPI PP  8..89*****
MYG_MOUSE      g--LSDGEWQLVLNVWGKVEADLAGHGQEVLI GLFKTHPETLDKFDKFKN
#=GR MYG_MOUSE PP  8..89*****
MYG_MUSAN      v-----DWEKVN SVWSAVESDLTAIGQNILRLRFEQY PESQNHFPKFKN
...

```

and so on.

Notice those PP annotation lines. That's posterior probability annotation, as in the single sequence alignments that `hmmsearch` and `hmmalign` showed. This essentially represents the confidence that each residue is aligned where it should be.

`hmmalign` currently has a “feature” that we’re aware of. Recall that HMMER3 only does local alignments. Here, we know that we’ve provided full length globin sequences, and `globins4` is a full length globin model. We’d probably like `hmmalign` to produce a global alignment. It can’t currently do that. If it doesn’t quite manage to extend its local alignment to the full length of a target globin sequence, you’ll get a weird-looking effect, as the nonmatching termini are pulled out to the left or right. For example, look at the N-terminal `g` in `MYG_HORSE` above. H3 is about 80% confident that this residue is nonhomologous, though any sensible person would align it into the first globin consensus column.

Look at the end of that first block of Stockholm alignment, where you’ll see:

```

...
HBBL_RANCA      v-HWTAEEKAVINSVWQKV--DVEQDGHEALTRLFIVYPWTQRYFSTFGD
#=GR HBBL_RANCA PP  6.6799*****
HBB2_TRICR      .VHLTAEDRKEIAAILGKV--NVDSLGGQCLARLIVVNPWSRRYFHDFGD
#=GR HBB2_TRICR PP  .69*****
#=GC PP_cons     .679*****
#=GC RF         .xxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxxx

```

The `#=GC PP_cons` line is Stockholm-format *consensus posterior probability* annotation for the entire column. It’s calculated simply as the arithmetic mean of the per-residue posterior probabilities in that column. This should prove useful in phylogenetic inference applications, for example, where it’s common to mask away nonconfidently aligned columns of a multiple alignment. The `PP_cons` line provides an objective measure of the confidence assigned to each column.

The `#=GC RF` line is Stockholm-format *reference coordinate annotation*, with an x marking each column that the profile considered to be consensus.

Single sequence queries using phmmer

The `phmmer` program is for searching a single sequence query against a sequence database, much as BLASTP or FASTA would do. `phmmer` works essentially just like `hmmsearch` does, except you provide a query sequence instead of a query profile HMM.

Internally, HMMER builds a profile HMM from your single query sequence, using a simple position-independent scoring system (BLOSUM62 scores converted to probabilities, plus a gap-open and gap-extend probability).

The file `tutorial/HBB_HUMAN` is a FASTA file containing the human β -globin sequence as an example query. If you have a sequence database such as `uniprot_sprot.fasta`, make that your target database; otherwise, use `tutorial/globins45.fa` as a small example:

```

> phmmer tutorial/HBB_HUMAN uniprot_sprot.fa
or
> phmmer tutorial/HBB_HUMAN tutorial/globins45.fa

```

Everything about the output is essentially as previously described for `hmmsearch`.

Iterative searches using jackhmmmer

The `jackhmmmer` program is for searching a single sequence query iteratively against a sequence database, much as `PSI-BLAST` would do.

The first round is identical to a `phmmer` search. All the matches that pass the inclusion thresholds are put in a multiple alignment. In the second (and subsequent) rounds, a profile is made from these results, and the database is searched again with the profile.

Iterations continue either until no new sequences are detected or the maximum number of iterations is reached. By default, the maximum number of iterations is 5; you can change this with the `-N` option.

Your original query sequence is always included in the multiple alignments, whether or not it appears in the database.⁵ The “consensus” columns assigned to each multiple alignment always correspond exactly to the residues of your query, so the coordinate system of every profile is always the same as the numbering of residues in your query sequence, 1..L for a sequence of length L.

Assuming you have Uniprot or something like it handy, here’s an example command line for a `jackhmmmer` search:

```
> jackhmmmer tutorial/HBB_HUMAN uniprot_sprot.fa
```

One difference from `phmmer` output you’ll notice is that `jackhmmmer` marks “new” sequences with a `+` and “lost” sequences with a `-`. New sequences are sequences that pass the inclusion threshold(s) in this round, but didn’t in the round before. Lost sequences are the opposite: sequences that passed the inclusion threshold(s) in the previous round, but have now fallen beneath (yet are still in the reported hits – it’s possible, though rare, to lose sequences utterly, if they no longer even pass the reporting threshold(s)). In the first round, everything above the inclusion thresholds is marked with a `+`, and nothing is marked with a `-`. For example, the top of this output looks like:

```
# jackhmmmer :: iteratively search a protein sequence against a protein database
# HMMER 3.0b3 (November 2009); http://hmmmer.org/
# Copyright (C) 2009 Howard Hughes Medical Institute.
# Freely distributed under the GNU General Public License (GPLv3).
# -----
# query sequence file:          HBB_HUMAN
# target sequence database:     uniprot_sprot.fasta
# per-seq hits tabular output:  hbb-jack.tbl
# per-dom hits tabular output:  hbb-jack.domtbl
# -----

Query:          HBB_HUMAN  [L=146]
Description:    Human beta hemoglobin.

Scores for complete sequences (score includes all domains):
--- full sequence ---  --- best 1 domain ---  -#dom-
E-value  score  bias    E-value  score  bias    exp  N  Sequence                Description
+  2.3e-98  331.4  0.0    2.5e-98  331.2  0.0    1.0  1  sp|P68871|HBB_HUMAN      Hemoglobin subunit beta OS=Homo sapien
+  2.3e-98  331.4  0.0    2.5e-98  331.2  0.0    1.0  1  sp|P68872|HBB_PANPA      Hemoglobin subunit beta OS=Pan paniscu
+  2.3e-98  331.4  0.0    2.5e-98  331.2  0.0    1.0  1  sp|P68873|HBB_PANTR      Hemoglobin subunit beta OS=Pan troglod
+  9.1e-98  329.4  0.0        1e-97  329.3  0.0    1.0  1  sp|P02024|HBB_GORGO      Hemoglobin subunit beta OS=Gorilla gor
+  2e-96  325.1  0.0    2.2e-96  324.9  0.0    1.0  1  sp|P02025|HBB_HYLLA      Hemoglobin subunit beta OS=Hylobates l
+  2e-95  321.8  0.0    2.2e-95  321.7  0.0    1.0  1  sp|P02032|HBB_SEMEN      Hemoglobin subunit beta OS=Semnopithec
...
```

That continues until the inclusion threshold is reached, at which point you see a tagline “inclusion threshold” indicating where the threshold was set:

```
+  0.00076  24.1  0.0    0.00083  24.0  0.0    1.0  1  sp|P02180|MYG_BALPH      Myoglobin OS=Balaenoptera physalus GN=
+  0.00087  23.9  0.0    0.0009  23.9  0.0    1.0  1  sp|P02148|MYG_PONPY      Myoglobin OS=Pongo pygmaeus GN=MB PE=1
```

⁵If *it* is in the database, it will almost certainly be included in the internal multiple alignment twice, once because it’s the query and once because it’s a significant database match to itself. This redundancy won’t screw up the alignment, because sequences are downweighted for redundancy anyway.

```

----- inclusion threshold -----
0.0013  23.3  0.3    0.021  19.4  0.2    2.1  1  sp|P81044|HBAZ_MACEU  Hemoglobin subunit zeta (Fragments) OS
0.0021  22.7  0.0    0.0022  22.6  0.0    1.0  1  sp|P02182|MYG_ZIPCA   Myoglobin OS=Ziphilus cavirostris GN=MB

```

The domain output and search statistics are then shown just as in `phmmer`. At the end of this first iteration, you'll see some output that starts with `@@` (this is a simple tag that lets you search through the file to find the end of one iteration and the beginning of another):

```

@@ New targets included: 894
@@ New alignment includes: 895 subseqs (was 1), including original query
@@ Continuing to next round.

@@
@@ Round: 2
@@ Included in MSA: 895 subsequences (query + 894 subseqs from 894 targets)
@@ Model size: 146 positions
@@

```

This (obviously) is telling you that the new alignment contains 895 sequences, your query plus 894 significant matches. For round two, it's built a new model from this alignment. Now for round two, it fires off what's essentially an `hmmsearch` of the target database with this new model:

```

Scores for complete sequences (score includes all domains):
--- full sequence --- --- best 1 domain --- -#dom-
E-value  score  bias  E-value  score  bias  exp  N  Sequence  Description
-----  -
1.5e-67  231.0  0.2    1.7e-67  230.8  0.1    1.0  1  sp|P02055|HBB_MELME  Hemoglobin subunit beta OS=Meles meles
2.3e-67  230.4  0.4    2.6e-67  230.2  0.3    1.0  1  sp|P81042|HBE_MACEU  Hemoglobin subunit epsilon OS=Macropus
2.7e-67  230.2  0.3    2.9e-67  230.0  0.2    1.0  1  sp|P15449|HBB_MELCA  Hemoglobin subunit beta OS=Mellivora c
3.3e-67  229.9  0.2    3.7e-67  229.7  0.2    1.0  1  sp|P68046|HBB_ODORO  Hemoglobin subunit beta OS=Odobenus ro
...

```

If you skim down through this output, you'll start seeing newly included sequences marked with `+`'s, such as:

```

...
+ 9.8e-30  108.3  0.0    1.1e-29  108.2  0.0    1.0  1  sp|P87497|MYG_CHIRA  Myoglobin OS=Chionodraco rastrospinosu
+ 1.4e-29  107.8  0.3    1.5e-29  107.7  0.2    1.0  1  sp|P14399|MYG_MUSAN  Myoglobin OS=Mustelus antarcticus GN=m
+ 1.5e-29  107.7  0.3    2e-29  107.3  0.2    1.0  1  sp|P80017|GLBD_CAUAR  Globin D, coelomic OS=Caudina arenicol
+ 3e-29  106.8  0.0    3.3e-29  106.6  0.0    1.0  1  sp|P02022|HBAM_RANCA  Hemoglobin heart muscle subunit alpha
+ 4e-29  106.3  0.0    4.4e-29  106.2  0.0    1.0  1  sp|Q9DEP0|MYG_CRYAN  Myoglobin OS=Cryodraco antarcticus GN=
+ 9.3e-29  105.2  0.2    1e-28  105.0  0.1    1.0  1  sp|P14397|MYG_GALGA  Myoglobin OS=Galeorhinus galeus GN=mb
+ 1.3e-28  104.7  0.0    1.6e-28  104.4  0.0    1.0  1  sp|P0C227|GLB_NERAL  Globin OS=Nerita albicilla PE=1 SV=1
+ 2e-28  104.1  0.0    2.4e-28  103.8  0.0    1.0  1  sp|P09106|HBAT_PAPAN  Hemoglobin subunit theta-1 OS=Papio an
+ 2.8e-28  103.6  0.1    3.1e-28  103.5  0.1    1.0  1  sp|P14398|MYG_GALJA  Myoglobin OS=Galeorhinus japonicus GN=
+ 7.9e-26  95.7  0.0    8.8e-26  95.5  0.0    1.0  1  sp|P23216|GLBP1_GLYDI  Globin, major polymeric component P1 O
...

```

It's unusual to see sequences get lost (and marked with `-`), but it can happen; it doesn't happen in this globin example.

After round 2, many more globin sequences have been found:

```

@@ New targets included: 167
@@ New alignment includes: 1064 subseqs (was 895), including original query
@@ Continuing to next round.

@@
@@ Round: 3
@@ Included in MSA: 1064 subsequences (query + 1063 subseqs from 1061 targets)
@@ Model size: 146 positions
@@

```

Because new sequences were included, it keeps going to round three, and then again to round four, then again to round five. After round five (where this example has found 1113 included hits in the database), the search ends quietly because there's a default maximum of five iterations, and you get:

```

@@ New targets included: 1
@@ New alignment includes: 1114 subseqs (was 1113), including original query
//

```

That `//` marks the end of the results for one query.

4 File formats

HMMER profile HMM files

The file `tutorial/fn3.hmm` gives an example of a HMMER3 ASCII save file. An abridged version is shown here, where (...) mark deletions made for clarity and space:

```
HMMER3/b [3.0b2 | June 2009]
NAME fn3
ACC PF00041.12
DESC Fibronectin type III domain
LENG 86
ALPH amino
RF no
CS yes
MAP yes
DATE Mon Jun 15 09:39:41 2009
NSEQ 106
EFFN 11.415833
CKSUM 72638555
GA 8.00 7.20
TC 8.00 7.20
NC 7.90 7.90
STATS LOCAL MSV -9.4043 0.71847
STATS LOCAL VITERBI -9.7737 0.71847
STATS LOCAL FORWARD -3.8341 0.71847
HMM
      A          C          D          E          F          G          H          I          (...)  Y
      m->m      m->i      m->d      i->m      i->i      d->m      d->d
COMPO 2.70271 4.89246 3.02314 2.64362 3.59817 2.82566 3.74147 3.08574 (...) 3.22607
      2.68618 4.42225 2.77519 2.73123 3.46354 2.40513 3.72494 3.29354 (...) 3.61503
      0.00338 6.08833 6.81068 0.61958 0.77255 0.00000 *
      1 3.16986 5.21447 4.52134 3.29953 4.34285 4.18764 4.30886 3.35801 (...) 3.93889 1 -
      2.68629 4.42236 2.77530 2.73088 3.46365 2.40512 3.72505 3.29365 (...) 3.61514
      0.09796 2.38361 6.81068 0.10064 2.34607 0.48576 0.95510
      2 2.70230 5.97353 2.24744 2.62947 5.31433 2.60356 4.43584 4.79731 (...) 4.25623 3 -
      2.68618 4.42225 2.77519 2.73123 3.46354 2.40513 3.72494 3.29354 (...) 3.61503
      0.00338 6.08833 6.81068 0.61958 0.77255 0.48576 0.95510
(...)
      85 2.48488 5.72055 3.87501 1.97538 3.04853 3.48010 4.51877 3.51898 (...) 3.43366 120 - B
      2.68618 4.42225 2.77519 2.73123 3.46354 2.40513 3.72494 3.29354 (...) 3.61503
      0.00338 6.08833 6.81068 0.61958 0.77255 0.48576 0.95510
      86 3.03720 5.94099 3.75455 2.96917 5.26587 2.91682 3.66571 4.11840 (...) 4.99111 121 - E
      2.68618 4.42225 2.77519 2.73123 3.46354 2.40513 3.72494 3.29354 (...) 3.61503
      0.00227 6.08723 * 0.61958 0.77255 0.00000 *
//
```

An HMM file consists of one or more HMMs. Each HMM starts with the identifier `HMMER3/b` and ends with `//` on a line by itself. The identifier allows backward compatibility as the HMMER software evolves: it tells the parser this file is from HMMER3's save file format version b. (The 3/a format was used in alpha test versions of HMMER3.) The closing `//` allows multiple HMMs to be concatenated.

The format is divided into two regions. The first region contains textual information and miscellaneous parameters in a roughly tag-value scheme. This section ends with a line beginning with the keyword `HMM`. The second region is a tabular, whitespace-limited format for the main model parameters.

All probability parameters are all stored as negative natural log probabilities with five digits of precision to the right of the decimal point, rounded. For example, a probability of 0.25 is stored as $-\log 0.25 = 1.38629$. The special case of a zero probability is stored as `*`.

Spacing is arranged for human readability, but the parser only cares that fields are separated by at least one space character.

A more detailed description of the format follows.

header section

The header section is parsed line by line in a tag/value format. Each line type is either **mandatory** or **optional** as indicated.

HMMER3/b Unique identifier for the save file format version; the `/b` means that this is HMMER3 HMM file format version b. When HMMER3 changes its save file format, the revision code advances. This way, parsers may easily remain backwards compatible. The remainder of the line after the `HMMER3/b` tag is free text that is ignored by the parser. HMMER currently writes its version number and release date in brackets here, e.g. `[3.0b2 | June 2009]` in this example. **Mandatory.**

- NAME** <s> Model name; <s> is a single word containing no spaces or tabs. The name is normally picked up from the `#=GF ID` line from a Stockholm alignment file. If this is not present, the name is created from the name of the alignment file by removing any file type suffix. For example, an otherwise nameless HMM built from the alignment file `rrm.slx` would be named `rrm`. **Mandatory**.
- ACC** <s> Accession number; <s> is a one-word accession number. This is picked up from the `#=GF AC` line in a Stockholm format alignment. **Optional**.
- DESC** <s> Description line; <s> is a one-line free text description. This is picked up from the `#=GF DE` line in a Stockholm alignment file. **Optional**.
- LENG** <d> Model length; <d>, a positive nonzero integer, is the number of match states in the model. **Mandatory**.
- ALPH** <s> Symbol alphabet type. For biosequence analysis models, <s> is `amino`, `DNA`, or `RNA` (case insensitive). There are also other accepted alphabets for purposes beyond biosequence analysis, including `coins`, `dice`, and `custom`. This determines the symbol alphabet and the size of the symbol emission probability distributions. If `amino`, the alphabet size K is set to 20 and the symbol alphabet to “ACDEFGHIKLMNPQRSTVWY” (alphabetic order); if `DNA`, the alphabet size K is set to 4 and the symbol alphabet to “ACGT”; if `RNA`, the alphabet size K is set to 4 and the symbol alphabet to “ACGU”. **Mandatory**.
- RF** <s> Reference annotation flag; <s> is either `no` or `yes` (case insensitive). If `yes`, the reference annotation character field for each match state in the main model (see below) is valid; if `no`, these characters are ignored. Reference column annotation is picked up from a Stockholm alignment file’s `#=GC RF` line. It is propagated to alignment outputs, and also may optionally be used to define consensus match columns in profile HMM construction. **Optional**; assumed to be `no` if not present.
- CS** <s> Consensus structure annotation flag; <s> is either `no` or `yes` (case insensitive). If `yes`, the consensus structure character field for each match state in the main model (see below) is valid; if `no` these characters are ignored. Consensus structure annotation is picked up from a Stockholm file’s `#=GC SS_cons` line, and propagated to alignment displays. **Optional**; assumed to be `no` if not present.
- MAP** <s> Map annotation flag; <s> is either `no` or `yes` (case insensitive). If set to `yes`, the map annotation field in the main model (see below) is valid; if `no`, that field will be ignored. The HMM/alignment map annotates each match state with the index of the alignment column from which it came. It can be used for quickly mapping any subsequent HMM alignment back to the original multiple alignment, via the model. **Optional**; assumed to be `no` if not present.
- DATE** <s> Date the model was constructed; <s> is a free text date string. This field is only used for logging purposes.¹ **Optional**.
- COM** [<n>] <s> Command line log; <n> counts command line numbers, and <s> is a one-line command. There may be more than one `COM` line per save file, each numbered starting from $n = 1$. These lines record every HMMER command that modified the save file. This helps us reproducibly and automatically log how Pfam models have been constructed, for example. **Optional**.

¹HMMER does not use dates for any purpose other than human-readable annotation, so it is no more prone than you are to Y2K, Y2038, or any other date-related eschatology.

- NSEQ** <d> Sequence number; <d> is a nonzero positive integer, the number of sequences that the HMM was trained on. This field is only used for logging purposes. **Optional.**
- EFFN** <f> Effective sequence number; <f> is a nonzero positive real, the effective total number of sequences determined by `hmmbuild` during sequence weighting, for combining observed counts with Dirichlet prior information in parameterizing the model. This field is only used for logging purposes. **Optional.**
- CKSUM** <d> Training alignment checksum; <d> is a nonnegative unsigned 32-bit integer. This number is calculated from the training sequence data, and used in conjunction with the alignment map information to verify that a given alignment is indeed the alignment that the map is for. **Optional.**
- GA** <f> <f> Pfam gathering thresholds GA1 and GA2. See Pfam documentation of GA lines. **Optional.**
- TC** <f> <f> Pfam trusted cutoffs TC1 and TC2. See Pfam documentation of TC lines. **Optional.**
- NC** <f> <f> Pfam noise cutoffs NC1 and NC2. See Pfam documentation of NC lines. **Optional.**
- STATS** <s1> <s2> <f1> <f2> Statistical parameters needed for E-value calculations. <s1> is the model's alignment mode configuration: currently only `LOCAL` is recognized. <s2> is the name of the score distribution: currently `MSV`, `VITERBI`, and `FORWARD` are recognized. <f1> and <f2> are two real-valued parameters controlling location and slope of each distribution, respectively; μ and λ for Gumbel distributions for MSV and Viterbi scores, and τ and λ for exponential tails for Forward scores. λ values must be positive. All three lines or none of them must be present: when all three are present, the model is considered to be calibrated for E-value statistics. **Optional.**
- HMM** Flags the start of the main model section. Solely for human readability of the tabular model data, the symbol alphabet is shown on the `HMM` line, aligned to the fields of the match and insert symbol emission distributions in the main model below. The next line is also for human readability, providing column headers for the state transition probability fields in the main model section that follows. Though unparsed after the `HMM` tag, the presence of two header lines is **mandatory**: the parser always skips the line after the `HMM` tag line.
- COMPO** <f>*K The first line in the main model section may be an optional line starting with `COMPO`: these are the model's overall average match state emission probabilities, which are used as a background residue composition in the "filter null" model. The *K* fields on this line are log probabilities for each residue in the appropriate biosequence alphabet's order. **Optional.**

main model section

All the remaining fields are **mandatory**.

The first two lines in the main model section are atypical.² They contain information for the core model's BEGIN node. This is stored as model node 0, and match state 0 is treated as the BEGIN state. The begin state is mute, so there are no match emission probabilities. The first line is the insert 0 emissions. The second line contains the transitions from the begin state and insert state 0. These seven numbers are:

²That is, the first two lines after the optional `COMPO` line. Don't be confused by the presence of an optional `COMPO` line here. The `COMPO` line is placed in the model section, below the residue column headers, because it's an array of numbers much like residue scores, but it's not really part of the model.

$B \rightarrow M_1, B \rightarrow I_0, B \rightarrow D_1; I_0 \rightarrow M_1, I_0 \rightarrow I_0$; then a 0.0 and a '*', because by convention, nonexistent transitions from the nonexistent delete state 0 are set to $\log 1 = 0$ and $\log 0 = -\infty = '*'$.

The remainder of the model has three lines per node, for M nodes (where M is the number of match states, as given by the `LENG` line). These three lines are (K is the alphabet size in residues):

Match emission line The first field is the node number ($1 \dots M$). The parser verifies this number as a consistency check (it expects the nodes to come in order). The next K numbers for match emissions, one per symbol, in alphabetic order.

The next field is the `MAP` annotation for this node. If `MAP` was `yes` in the header, then this is an integer, representing the alignment column index for this match state ($1..alen$); otherwise, this field is '-'.

The next field is the `RF` annotation for this node. If `RF` was `yes` in the header, then this is a single character, representing the reference annotation for this match state; otherwise, this field is '-'.

The next field is the `CS` annotation for this node. If `CS` was `yes`, then this is a single character, representing the consensus structure at this match state; otherwise this field is '-'.

Insert emission line The K fields on this line are the insert emission scores, one per symbol, in alphabetic order.

State transition line The seven fields on this line are the transitions for node k , in the order shown by the transition header line: $M_k \rightarrow M_{k+1}, I_k, D_{k+1}; I_k \rightarrow M_{k+1}, I_k; D_k \rightarrow M_{k+1}, D_{k+1}$.

For transitions from the final node M , match state $M + 1$ is interpreted as the END state E , and there is no delete state $M + 1$; therefore the final $M_k \rightarrow D_{k+1}$ and $D_k \rightarrow D_{k+1}$ transitions are always * (zero probability), and the final $D_k \rightarrow M_{k+1}$ transition is always 0.0 (probability 1.0).

Finally, the last line of the format is the "//" record separator.

Stockholm, the recommended multiple sequence alignment format

The Pfam and Rfam Consortia have developed a multiple sequence alignment format called "Stockholm format" that allows rich and extensible annotation.

Most popular multiple alignment file formats can be changed into a minimal Stockholm format file just by adding a Stockholm header line and a trailing // terminator:

```
# STOCKHOLM 1.0

seq1 ACDEF...GHIKL
seq2 ACDEF...GHIKL
seq3 ...EFMNRGHIKL

seq1 MNPQTVWY
seq2 MNPQTVWY
seq3 MNPQT...
//
```

The first line in the file must be `# STOCKHOLM 1.x`, where x is a minor version number for the format specification (and which currently has no effect on my parsers). This line allows a parser to instantly identify the file format.

In the alignment, each line contains a name, followed by the aligned sequence. A dash, period, underscore, or tilde (but not whitespace) denotes a gap. If the alignment is too long to fit on one line, the alignment may be split into multiple blocks, with blocks separated by blank lines. The number of sequences, their order, and their names must be the same in every block. Within a given block, each (sub)sequence

(and any associated `#=GR` and `#=GC` markup, see below) is of equal length, called the *block length*. Block lengths may differ from block to block. The block length must be at least one residue, and there is no maximum.

Other blank lines are ignored. You can add comments anywhere to the file (even within a block) on lines starting with a `#`.

All other annotation is added using a tag/value comment style. The tag/value format is inherently extensible, and readily made backwards-compatible; unrecognized tags will simply be ignored. Extra annotation includes consensus and individual RNA or protein secondary structure, sequence weights, a reference coordinate system for the columns, and database source information including name, accession number, and coordinates (for subsequences extracted from a longer source sequence) See below for details.

syntax of Stockholm markup

There are four types of Stockholm markup annotation, for per-file, per-sequence, per-column, and per-residue annotation:

`#=GF <tag> <s>` Per-file annotation. `<s>` is a free format text line of annotation type `<tag>`. For example, `#=GF DATE April 1, 2000`. Can occur anywhere in the file, but usually all the `#=GF` markups occur in a header.

`#=GS <seqname> <tag> <s>` Per-sequence annotation. `<s>` is a free format text line of annotation type `tag` associated with the sequence named `<seqname>`. For example, `#=GS seq1 SPECIES.SOURCE Caenorhabditis elegans`. Can occur anywhere in the file, but in single-block formats (e.g. the Pfam distribution) will typically follow on the line after the sequence itself, and in multi-block formats (e.g. HMMER output), will typically occur in the header preceding the alignment but following the `#=GF` annotation.

`#=GC <tag> <..s..>` Per-column annotation. `<..s..>` is an aligned text line of annotation type `<tag>`. `#=GC` lines are associated with a sequence alignment block; `<..s..>` is aligned to the residues in the alignment block, and has the same length as the rest of the block. Typically `#=GC` lines are placed at the end of each block.

`#=GR <seqname> <tag> <..s..>` Per-residue annotation. `<..s..>` is an aligned text line of annotation type `<tag>`, associated with the sequence named `<seqname>`. `#=GR` lines are associated with one sequence in a sequence alignment block; `<..s..>` is aligned to the residues in that sequence, and has the same length as the rest of the block. Typically `#=GR` lines are placed immediately following the aligned sequence they annotate.

semantics of Stockholm markup

Any Stockholm parser will accept syntactically correct files, but is not obligated to do anything with the markup lines. It is up to the application whether it will attempt to interpret the meaning (the semantics) of the markup in a useful way. At the two extremes are the Belvu alignment viewer and the HMMER profile hidden Markov model software package.

Belvu simply reads Stockholm markup and displays it, without trying to interpret it at all. The tag types (`#=GF`, etc.) are sufficient to tell Belvu how to display the markup: whether it is attached to the whole file, sequences, columns, or residues.

HMMER uses Stockholm markup to pick up a variety of information from the Pfam multiple alignment database. The Pfam consortium therefore agrees on additional syntax for certain tag types, so HMMER can parse some markups for useful information. This additional syntax is imposed by Pfam, HMMER, and

other software of mine, not by Stockholm format per se. You can think of Stockholm as akin to XML, and what my software reads as akin to an XML DTD, if you're into that sort of structured data format lingo.

The Stockholm markup tags that are parsed semantically by my software are as follows:

recognized #=GF annotations

- ID** <s> Identifier. <s> is a name for the alignment; e.g. "rrm". One word. Unique in file.
- AC** <s> Accession. <s> is a unique accession number for the alignment; e.g. "PF00001". Used by the Pfam database, for instance. Often a alphabetical prefix indicating the database (e.g. "PF") followed by a unique numerical accession. One word. Unique in file.
- DE** <s> Description. <s> is a free format line giving a description of the alignment; e.g. "RNA recognition motif proteins". One line. Unique in file.
- AU** <s> Author. <s> is a free format line listing the authors responsible for an alignment; e.g. "Bateman A". One line. Unique in file.
- GA** <f> <f> Gathering thresholds. Two real numbers giving HMMER bit score per-sequence and per-domain cutoffs used in gathering the members of Pfam full alignments.
- NC** <f> <f> Noise cutoffs. Two real numbers giving HMMER bit score per-sequence and per-domain cutoffs, set according to the highest scores seen for unrelated sequences when gathering members of Pfam full alignments.
- TC** <f> <f> Trusted cutoffs. Two real numbers giving HMMER bit score per-sequence and per-domain cutoffs, set according to the lowest scores seen for true homologous sequences that were above the GA gathering thresholds, when gathering members of Pfam full alignments.

recognized #=GS annotations

- WT** <f> Weight. <f> is a positive real number giving the relative weight for a sequence, usually used to compensate for biased representation by downweighting similar sequences. Usually the weights average 1.0 (e.g. the weights sum to the number of sequences in the alignment) but this is not required. Either every sequence must have a weight annotated, or none of them can.
- AC** <s> Accession. <s> is a database accession number for this sequence. (Compare the #=GF AC markup, which gives an accession for the whole alignment.) One word.
- DE** <s> Description. <s> is one line giving a description for this sequence. (Compare the #=GF DE markup, which gives a description for the whole alignment.)

recognized #=GC annotations

- RF** Reference line. Any character is accepted as a markup for a column. The intent is to allow labeling the columns with some sort of mark.
- SS.cons** Secondary structure consensus. For protein alignments, DSSP codes or gaps are accepted as markup: [HGIEBTSCX.-_], where H is alpha helix, G is 3/10-helix, I is p-helix, E is extended strand, B is a residue in an isolated b-bridge, T is a turn, S is a bend, C is a random coil or loop, and X is unknown (for instance, a residue that was not resolved in a crystal structure).
- SA.cons** Surface accessibility consensus. 0-9, gap symbols, or X are accepted as markup. 0 means <10% accessible residue surface area, 1 means <20%, 9 means <100%, etc. X means unknown structure.

recognized #-GR annotations

ss Secondary structure consensus. See #-GC SS_cons above.

sa Surface accessibility consensus. See #-GC SA_cons above.

pp Posterior probability for an aligned residue. This represents the probability that this residue is assigned to the HMM state corresponding to this alignment column, as opposed to some other state. (Note that a residue can be confidently *unaligned*: a residue in an insert state or flanking N or C state may have high posterior probability.) The posterior probability is encoded as 11 possible characters 0-9*: $0.0 \leq p < 0.05$ is coded as 0, $0.05 \leq p < 0.15$ is coded as 1, (... and so on ...), $0.85 \leq p < 0.95$ is coded as 9, and $0.95 \leq p \leq 1.0$ is coded as '*'. Gap characters appear in the PP line where no residue has been assigned.

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